

Definition of the Labile Capping Bond Effect in Lanthanide Complexes

Aurora Rodríguez-Rodríguez^{a,b}, Martín Regueiro-Figueroa^a, David Esteban-Gómez^a, Teresa Rodríguez-Blas^a, Véronique Patinec^b, Raphaël Tripier^b, Gyula Tircsó^c, Fabio Carniato^d, Mauro Botta^d and Carlos Platas-Iglesias^a

^a Centro de Investigaciones Científicas Avanzadas (CICA), Departamento de Química Fundamental, Facultad de Ciencias, Universidade da Coruña, A Coruña, Galicia, Spain

^b Université de Bretagne Occidentale, UMR-CNRS 6521, UFR des Sciences et Techniques, Brest Cedex 3, France

^c Department of Inorganic and Analytical Chemistry, University of Debrecen, Debrecen, Egyetem tér 1, Hungary

^d Dipartimento di Scienze e Innovazione Tecnologica, Università del Piemonte Orientale "A. Avogadro", Alessandria, Italy

Chemistry - A European Journal Volume 23, Issue 5, pages 1110–1117, January 23, 2017

Issue online: 25 January 2017, Version of record online: 19 December 2016, Accepted manuscript online: 8 November 2016, Manuscript Received: 16 September 2016

This is the peer reviewed version of the following article:

Rodríguez-Rodríguez, A., Regueiro-Figueroa, M., Esteban-Gómez, D., Rodríguez-Blas, T., Patinec, V., Tripier, R., Tircsó, G., Carniato, F., Botta, M., Platas-Iglesias, C. (2017), Definition of the Labile Capping Bond Effect in Lanthanide Complexes. *Chem. Eur. J.*, 23: 1110-1117.

which has been published in final form at <https://doi.org/10.1002/chem.201604390>. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Self-Archiving.

Abstract

Two macrocyclic ligands containing a cyclen unit, a methyl group, a picolinate arm, and two acetate pendant arms attached to two nitrogen atoms of the macrocycle either in *trans* (1,7-H₃Medo2 ampa = 2,2'-(7-((6-carboxypyridin-2-yl)methyl)-10-methyl-1,4,7,10-tetraazacyclododecane-1,4-diyl)diacetic acid) or in *cis* (1,4-H₃Medo2 ampa) positions are reported. These ligands provide eight-coordination to the Ln³⁺ ions, leaving a coordination position available for a water molecule that occupies a capping position in the twisted square antiprismatic polyhedron (1,4-H₃Medo2 ampa) or one of the positions of the square antiprism (1,7-H₃Medo2 ampa). The charge neutral [Gd(1,7-Medo2 ampa)] complex presents an unprecedentedly low water-exchange rate ($k_{\text{ex}}^{298}=8.8\times 10^3\text{ s}^{-1}$), whereas water exchange in [Gd(1,4-Medo2 ampa)] is three orders of magnitude faster ($k_{\text{ex}}^{298}=6.6\times 10^6\text{ s}^{-1}$). These results showcase the labile capping bond phenomenon: A ligand occupying a capping position is hindered by the environment and thus is intrinsically labile.

Keywords: density functional calculations; gadolinium; magnetic resonance imaging; NMR spectroscopy; water exchange

Introduction

Exchange reactions involving water molecules in the first and second solvation shells of metal complexes are of fundamental importance to understand the reactivity of metal ions in both chemical and biological

systems.^[1] The replacement of a coordinated water molecule by an entering ligand represents a key step in the formation of metal complexes in aqueous solutions^[2] and in many redox processes.^[3] The mean residence times of coordinated water molecules in aquated complexes spread over 20 orders of magnitude from approximately 300 years for $[\text{Ir}(\text{H}_2\text{O})_6]^{3+}$ to only approximately 200 ps for the extremely labile $[\text{Eu}(\text{H}_2\text{O})_7]^{2+}$ (at 25 °C).^[4, 5]

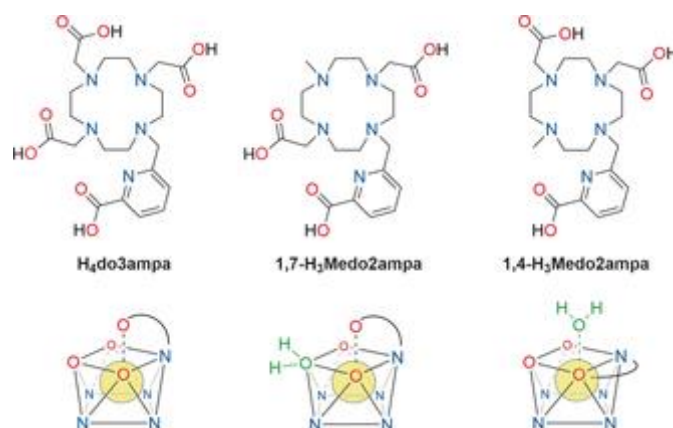
The water-exchange reactions in aquated lanthanide ions $[\text{Ln}(\text{H}_2\text{O})_q]^{3+}$ ($q=8$ or 9) were investigated by Merbach and co-workers using ^{17}O NMR techniques.^[6] Among the different Ln^{3+} ions, Gd^{3+} complexes have attracted particular attention during the last 20 years owing to their increasing use as contrast agents (CAs) in clinical and pre-clinical magnetic resonance imaging (MRI) procedures.^[7] CAs are paramagnetic compounds, often small Gd^{3+} chelates, that accelerate the relaxation rates of water molecules in the surrounding tissues.^[8] Water-exchange dynamics play a key role in determining the effectiveness of Gd^{3+} CAs, as exchange of the bound water molecule(s) should be sufficiently fast to attain optimal relaxivities.^[9] However, extremely slow water-exchange rates in Ln^{3+} complexes have been exploited to design CAs based on the chemical exchange saturation transfer (CEST) approach, which represent attractive alternatives to the classical Gd^{3+} -based agents.^[10] CEST agents based on Ln^{3+} ions typically contain a pool of exchangeable protons in intermediate-to-slow condition with the bulk water ($k_{\text{ex}} \leq \Delta\omega$, where $\Delta\omega$ is the frequency difference). Application of a presaturation pulse at the frequency of the exchangeable protons (i.e., a coordinated water molecule) induces the transfer of some saturated spins into the water pool, thereby attenuating the signal of bulk water.^[11]

The residence time of a water molecule in the inner coordination sphere of Gd^{3+} complexes (τ_m^{298}) expands over a range of approximately four orders of magnitude from the longest determined for dota-tetraamide (dota=1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid) derivatives ($\tau_m^{298}=8\text{--}20\ \mu\text{s}$)^[12] to the shortest measured for the aqua ion^[13] and a Gd^{3+} complex with an octadentate ligand containing phosphonate groups ($\tau_m^{298} \approx 1.3\text{--}1.4\ \text{ns}$).^[14] In the case of Eu^{3+} -based CEST agents, appropriate ligand modifications make it possible to attain residence times as long as 150–700 μs .^[15, 16]

Among the different factors that were identified to accelerate the water exchange of the coordinated water molecule in Gd^{3+} (or Eu^{3+}) complexes are 1) increasing the negative charge of the complex,^[17] and 2) increasing the steric compression around the water coordination site.^[18] Both effects facilitate the departure of the coordinated water molecule in a dissociative process, which is the most common mechanism responsible for the water-exchange reaction in nine-coordinate Gd^{3+} complexes.^[1] On the contrary, the inclusion of hydrophobic units around the water binding site minimizes hydrogen-bonding between the coordinated water and the second coordination sphere, which results in slower water-exchange rates.^[16, 19] The wider and more extensive number of studies focused on monoaqua nine-coordinated complexes. Many of these are derivatives of dota in which the water molecule occupies the apical position, capping the upper square face of the antiprism. A common and effective strategy for improving the effectiveness of a Gd^{3+} -chelate as MRI probe is to increase the hydration state from one ($q=1$) to two ($q=2$). In dota-like derivatives, this implies the presence of one water molecule in the axial position and of a second water molecule in an equatorial position, orthogonal to the first. A relevant yet still unanswered question concerning the water-exchange dynamics of these systems arises: Do the two water molecules show similar rates of exchange or do they behave independently?

Thus, we sought to design a pair of ligands for Ln^{3+} complexation having an identical number and type of donor atoms and providing complexes containing a coordinated water molecule either occupying a position in the coordination polyhedron or a capping position. This was accomplished by using as a starting point the $\text{H}_4\text{do3ampa}$ ligand ($\text{H}_4\text{do3ampa} = 2,2',2''\text{-(10-((6-carboxypyridin-2-yl)methyl)-1,4,7,10-tetraazacyclododecane-1,4,7-triyl)triacetic acid}$, Scheme 1),^[20] which was shown to form nine-coordinate complexes in solution in which the oxygen atom of the picolinate group occupies the capping position in a capped square antiprismatic (SAP) coordination polyhedron. We hypothesized that removing one of the carboxylate groups in *trans* position to the picolinate arm to afford 1,7- $\text{H}_3\text{Medo2ampa}$ should result in the

coordination of a water molecule in one of the square faces of the coordination polyhedron, with the capping position occupied by the oxygen atom of the picolinate group. In contrast, removal of a carboxylate group in *cis* position with respect to the pyridyl unit should allow picolinate to lean on the methyl group, permitting a water molecule to occupy a capping position. Thus, we report the synthesis of the 1,7-H₃Medo2 ampa and 1,4-H₃Medo2 ampa ligands, and demonstrate the labile capping bond effect with a detailed analysis of the water-exchange rates in the corresponding Gd³⁺ complexes. The discovery of this effect provides new strategies to modulate ligand-exchange rates in Ln³⁺ complexes, with great potential impact in the coordination chemistry of the 4f elements.



Scheme 1. Ligands discussed in the present work and their expected coordination mode after complexation with Gd³⁺.

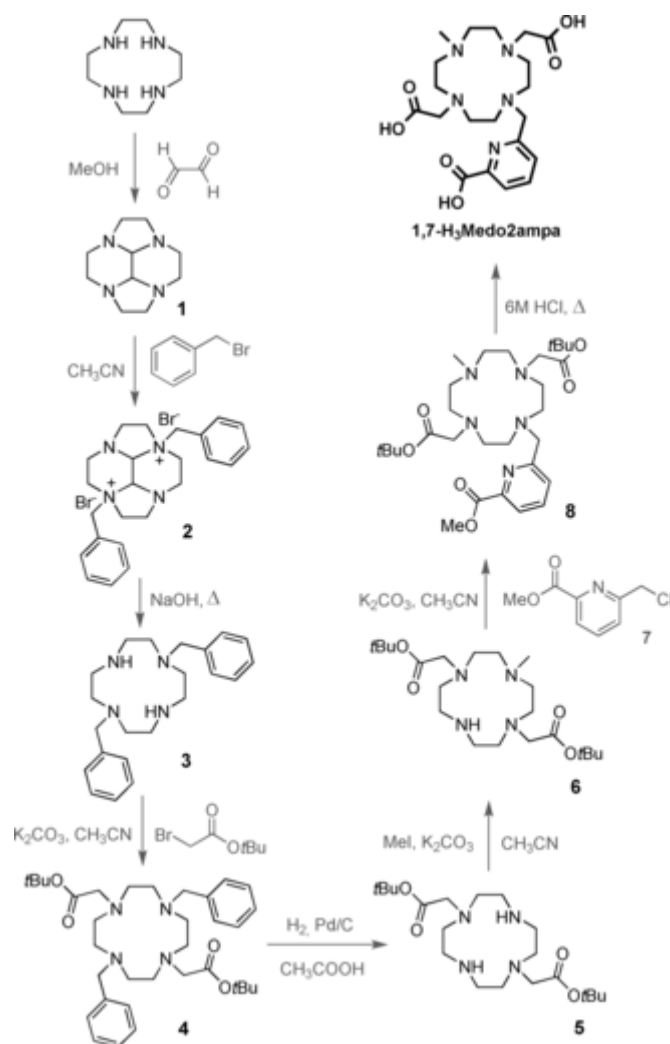
Results and Discussion

Synthesis of the ligands and metal complexes

The synthetic protocol used for the preparation of 1,7-H₃Medo2 ampa is shown in Scheme 2. Cyclen glyoxal (**1**) was obtained following the literature procedure by direct condensation of glyoxal with cyclen.^[21] The subsequent *trans*-alkylation of **1** with benzyl bromide afforded compound **2**,^[22] which was deprotected under basic conditions to furnish compound **3** with an overall yield of 93 % over the three steps. Reaction of **3** with *tert*-butyl bromoacetate in acetonitrile in the presence of K₂CO₃ afforded compound **4** in satisfactory yield (72 %). The benzyl groups of **4** were removed by Pd-catalyzed hydrogenolysis to yield compound **5** quantitatively. Monoalkylation of **5** with MeI to afford the key intermediate **6** was then achieved with 92 % yield. Careful selection of the reaction conditions was critical for the preparation of this compound in high yield. In particular, yields increased significantly if the reaction was performed at low temperature (0 °C), the concentration of precursor **5** in the reaction medium was not too high (<0.02 m), and MeI was added slowly to the solution of **5**. Alkylation of **6** with the 6-chloromethyl derivative **7**^[23] followed by deprotection of the methyl and *tert*-butyl ester groups provided the 1,7-H₃Medo2 ampa ligand, which was isolated as the hydrochloride salt with an overall yield of 50 % as calculated from cyclen (eight steps). The hydrochloride salt was then converted to the trifluoroacetate salt by treating the former with trifluoroacetic acid.

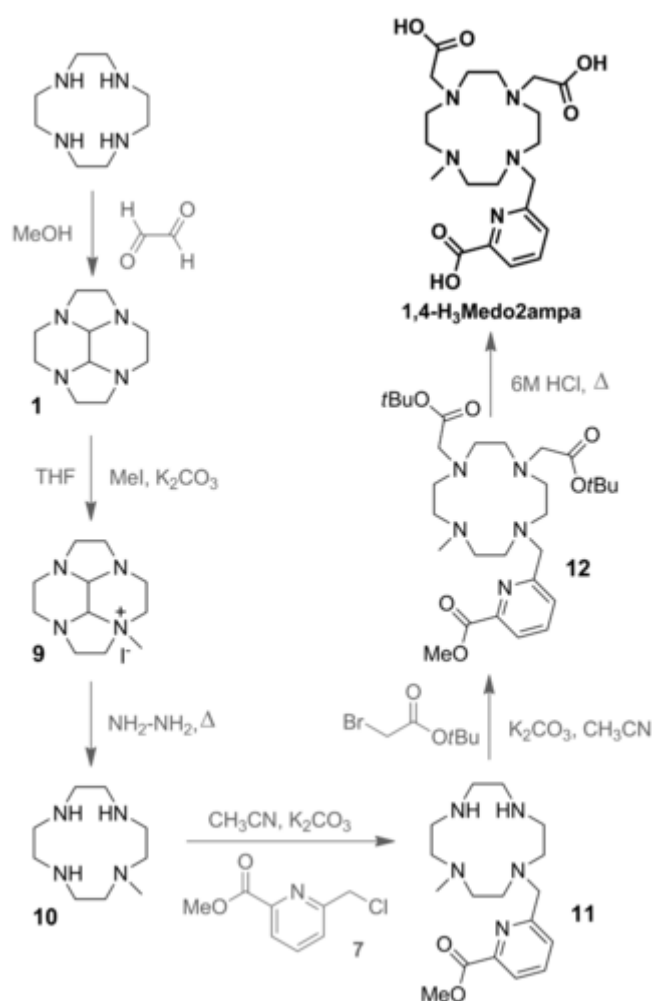
Reaction of 1,7-H₃Medo2 ampa with lanthanide triflates in the presence of an excess of triethylamine resulted in the formation of the charge-neutral complexes of formula [Ln(1,7-Medo2 ampa)(H₂O)] (Ln=La, Eu, Gd, Tb, Yb, or Lu), which were isolated in 85–90 % yield. The high-resolution mass spectra (HR-MS,

ESI⁺) show peaks resulting from the [Ln(1,7-HMedo2 ampa)]⁺ entities, thereby confirming the formation of the complexes (Figures S17–S22, Supporting Information).



Scheme 2. Synthesis of 1,7-H₃Medo2 ampa.

The synthesis of 1,4-H₃Medo2 ampa (Scheme 3) started with the conversion of **1** into the methylated derivative **9** by reaction with MeI (97%)^[22] followed by deprotection with hydrazine hydrate (87%). Subsequently, compound **10** was alkylated with compound **7**, affording the *cis* derivative **11**, which was used without further purification in the next step. Previous studies showed that the regioselective *cis* alkylation of cyclen depended both on the steric hindrance of electrophiles and the solvent system.^[24] The regioselective alkylation of **10** was achieved in acetonitrile at 0 °C. Finally, reaction of **11** with *tert*-butyl bromoacetate in acetonitrile in the presence of K₂CO₃ followed by deprotection of the methyl and *tert*-butyl ester groups with 6 m HCl provided the 1,4-H₃Medo2 ampa ligand in its hydrochloride salt form. The overall yield over the six steps required to prepare the ligand starting from cyclen was 31 %. The [Ln(1,7-Medo2 ampa)] complexes were prepared in aqueous solution by mixing equimolar amounts of the ligand and lanthanide chlorides or triflates, followed by adjustment of the pH value to 7.0. The HR-MS spectra (ESI⁺) confirm the formation of the complexes.



Scheme 3. Synthesis of 1,4-H₃Medo2 ampa.

Structural analysis

The ¹H NMR spectrum of the paramagnetic [Yb(1,7-Medo2 ampa)] complex (Figure 1) shows 25 of the 26 paramagnetically shifted signals expected for a single species with C₁ symmetry in solution. The most shifted axial protons of the cyclen ring are observed at 88.9, 82.3, 56.0, and 47.7 ppm. These values are very similar to those observed previously for the [Yb(do3 ampa)][−] complex (99.7, 78.8, 61.5, and 57.0 ppm).^[20] The latter compound was demonstrated to adopt an SAP structure in solution by analysis of the Yb³⁺-induced ¹H NMR shifts. Thus, we conclude that the [Yb(1,7-Medo2 ampa)] complex adopts a SAP structure in solution as well, which is confirmed by the relative energies of the two isomers obtained using DFT calculations (Supporting Information).

The ¹H NMR spectrum of the [Yb(1,4-Medo2 ampa)] complex (Figure 1) reveals the presence of two complex species in solution, which provide two sets of signals with different intensities. The most shifted axial proton of the minor species (ca. 30 %) is observed at 135.8 ppm, which is characteristic of a SAP coordination around the metal ions.^[25] Thus, the speciation of [Yb(1,4-Medo2 ampa)] in solution is dominated by the twisted-square antiprismatic (TSAP) isomer (ca. 70 %).

The number of water molecules coordinated to the Ln³⁺ ion (*q*) was assessed by measuring the lifetimes of the Eu³⁺(⁵D₀) and Tb³⁺(⁵D₄) excited states in solutions of the complexes in H₂O and D₂O (Table 1).^[26] The absorption spectra of the complexes recorded in H₂O solution show an absorption band with a maximum at 274 nm typical of the picolinate chromophore (Supporting Information).^[20] The emission spectra recorded

under excitation through the ligand bands (10^{-5} M, pH 7.0) show the ${}^5D_0 \rightarrow {}^7F_J$ ($J=0-4$) and ${}^5D_4 \rightarrow {}^7F_J$ ($J=3-6$) transitions characteristic of Eu^{3+} and Tb^{3+} , respectively (see the Supporting Information).

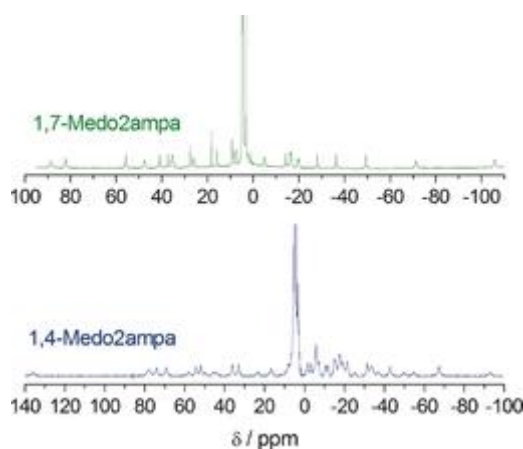


Figure 1. ${}^1\text{H}$ NMR spectra (300 MHz, D_2O , pH 7.0) of the Yb^{3+} complexes of 1,7-Medo2 ampa $^{3-}$ and 1,4-Medo2 ampa $^{3-}$.

Table 1. Luminescence lifetimes τ [ms] and hydration numbers q of the Eu^{3+} and Tb^{3+} complexes of 1,7-Medo2 ampa and 1,4-Medo2 ampa.

	Ln^{3+}	$\tau(\text{H}_2\text{O})$ [ms]	$\tau(\text{D}_2\text{O})$ [ms]	$\Delta k_{\text{obs}}^{[a]}$	$q^{[b]}$
1,7-Medo2 ampa	Eu^{3+}	0.569(3)	1.69(1)	1.17	1.1
	Tb^{3+}	1.793(5)	3.09(3)	0.23	0.9
1,4-Medo2 ampa	Eu^{3+}	0.584(2)	1.327(4)	0.96	0.9
	Tb^{3+}	2.39(1)	2.76(1)	0.06	0.0

[a] $\Delta k_{\text{obs}} = k_{\text{obs}}(\text{H}_2\text{O}) - k_{\text{obs}}(\text{D}_2\text{O})$; $k_{\text{obs}} = 1/\tau_{\text{obs}}$; [b] $q(\text{Eu}) = 1.2(\Delta k_{\text{obs}} - 0.25)$; $q(\text{Tb}) = 5.0(\Delta k_{\text{obs}} - 0.06)$.

The emission lifetimes of the $\text{Eu}^{3+}({}^5D_0)$ and $\text{Tb}^{3+}({}^5D_4)$ excited states of the complexes with 1,7-Medo2 ampa $^{3-}$ provide hydration numbers of $q=1.1$ and 0.9 according to the equation of Beeby.^[27] These results indicate that the Ln^{3+} complexes of 1,7-do2 ampa $^{3-}$ contain a water molecule in the inner-coordination sphere. This shows that the complexes are nine-coordinated in aqueous solution owing to the octadentate binding of the ligand and the presence of a coordinated water molecule. Application of the same methodology to the complexes with 1,4-do2 ampa $^{3-}$ provides a hydration number of 0.9 for Eu^{3+} and a hydration number of 0.0 for Tb^{3+} . These results suggest a rather abrupt change of the hydration number at the center of the lanthanide series.

To gain more insight into the structure of the complexes in aqueous solution, we performed DFT calculations at the TPSSh/LCRECP/6-31G(d) level.^[28-30] We have shown previously that a cluster/continuum approach

including explicitly two second-sphere water molecules provides a satisfactory description of the Gd–O_{water} distances and accurate ¹⁷O hyperfine coupling constants.^[31] The minimum energy conformation calculated for the SAP isomer of the [Gd(1,7-Medo2 ampa)(H₂O)]·2 H₂O system (Figure 2) indicates octadentate binding of the ligand to the Gd³⁺ ion. The basal plane of the antiprism is described by the four nitrogen atoms of the cyclen unit, whereas the upper plane is defined by the nitrogen atom of the pyridyl unit, two oxygen atoms of the acetate pendant arms, and a coordinated water molecule. The oxygen atom of the picolinate group occupies the capping position. The mean twist angle of the upper and lower square faces amounts to 36.7°.

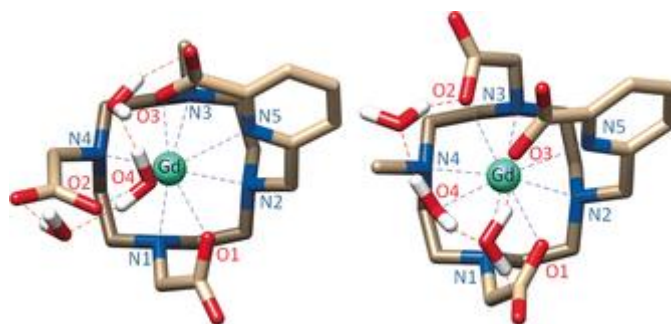


Figure 2. Structures of the [Gd(1,4-Medo2 ampa)(H₂O)]·2 H₂O (left) and [Gd(1,7-Medo2 ampa)(H₂O)]·2 H₂O (right) complexes optimized in aqueous solution at the TPSSh/LCRECP/6-31G(d) level.

Because the [Yb(1,4-Medo2 ampa)] complex exists in solution as a mixture of SAP and TSAP isomers we initially performed calculations on the [Gd(1,4-Medo2 ampa)(H₂O)] system. These calculations provided the SAP and TSAP isomers as minimum-energy conformations and predicted a rather small Gibbs energy difference between them, favoring the TSAP form by only approximately 2 kcal mol⁻¹. This energy reduces to 1.6 kcal mol⁻¹ for the Eu³⁺ complex. Thus, it is likely that both the SAP and TSAP isomers are present in solution in the case of the Gd³⁺ complex. The broad signals observed in the ¹H NMR spectrum of the Eu³⁺ complex are in line with this hypothesis. Most likely the SAP isomer contains a water molecule coordinated to the Ln³⁺ ion, whereas the sterically more crowded TSAP isomer does not contain a coordinated water molecule. The latter species is largely dominating in the case of the Tb³⁺ complex, thus resulting in a *q* value of 0.0 (Table 1).

The optimized structure of the SAP isomer of the [Gd(1,4-Medo2 ampa)(H₂O)]·2 H₂O complex reveals octadentate binding of the ligand, with the coordinated water molecule occupying the capping position above the mean plane delineated by the four donor atoms of the pendant arms. The calculated Gd–O_{water} distance (2.549 Å) is considerably longer than that calculated for [Gd(1,7-Medo2 ampa)(H₂O)]·2 H₂O (2.476 Å). These data reflect a weaker binding of the water molecule if it occupies a capping position in the coordination polyhedron. The calculated electron densities at the bond critical points (ρ_{BCP}) of the Gd–O_{water} bonds (0.041 and 0.034 a.u. for the complexes with 1,7-Medo2 ampa³⁻ and 1,4-Medo2 ampa³⁻, respectively) confirm that the coordinated water molecule is more tightly bound to the metal ion in the [Gd(1,7-Medo2 ampa)(H₂O)] complex.^[32]

Water-exchange rates of the coordinated water molecules

The water-exchange rates of the coordinated water molecules in the Gd³⁺ complexes were assessed by means of ¹⁷O NMR transverse relaxation rates and chemical shifts and ¹H relaxivity measurements. A combined analysis of the ¹H relaxivity and ¹⁷O NMR data is required for an accurate estimation of the water-exchange rates.^[33]

The relaxivities (r_{1p}) of an aqueous solution of the complexes were first assessed at 20 MHz and 25 °C (Figure 3). The relaxivity of [Gd(1,7-Medo2 ampa)] measured in the pH range 5.1–7.2 ($1.94 \text{ mm}^{-1} \text{ s}^{-1}$) is very low compared to the relaxivities of Gd^{3+} complexes containing one inner-sphere water molecule such as [Gd(dota)]⁻. Below pH 5.1, r_{1p} increases owing to the dissociation of the complex, with a relaxivity observed at pH < 2 that is very similar to that of $[\text{Gd}(\text{H}_2\text{O})]^{3+}$.^[13] Increasing the pH value above 7.2 results in a noticeable increase of r_{1p} , which reaches a value of $4.1 \text{ mm}^{-1} \text{ s}^{-1}$ at pH 11.1. This pH dependence of r_{1p} is characteristic of systems with very slow water-exchange rates around neutral pH value, so that the observed relaxivity is the result of the outer-sphere mechanism. Increasing the pH value favors an acceleration of the water exchange by OH^- catalysis of prototropic exchange.^[12, 34] The relaxivity of [Gd(1,4-Medo2 ampa)] at pH 7.0 (20 MHz and 25 °C) is $4.2 \text{ mm}^{-1} \text{ s}^{-1}$, and therefore considerably higher than that of the *trans* derivative. Again relaxivity increases below pH ≈ 4 owing to complex dissociation. Above pH 8 the relaxivity decreases slightly, likely resulting from the formation of hydroxo complexes.^[35]

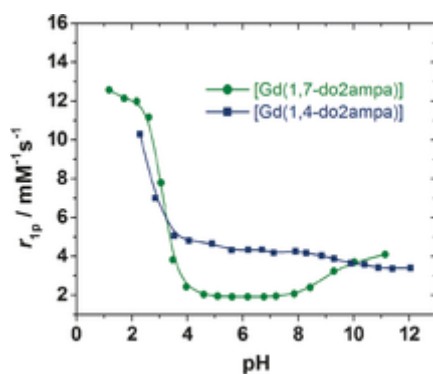


Figure 3. Plot of the ¹H relaxivities (20 MHz, 25 °C) of [Gd(1,7-Medo2 ampa)] and [Gd(1,4-Medo2 ampa)] as a function of pH value.

Additional insight into the exchange dynamics of the coordinated water molecules in these Gd^{3+} complexes was obtained by measuring the temperature dependence of r_{1p} (20 MHz, Figure 4). The relaxivity of [Gd(1,4-Medo2 ampa)] decreases with increasing temperature, a behavior typical of small chelates in which fast rotation of the complex in solution limits proton relaxivity. Conversely, the temperature dependence of r_{1p} measured for [Gd(1,7-Medo2 ampa)] at 20 MHz presents two distinct regions. From 5 to 30 °C the relaxivity is determined by the outer-sphere contribution, which increases as the temperature is reduced, whereas in the temperature range of 40–80 °C r_{1p} remains almost constant. This can be explained by an increasing contribution of the inner-sphere mechanism arising from a reduced τ_m (acceleration of the water-exchange rate), which is compensated by a decrease of the outer-sphere contribution and a faster rotation of the complex in solution at high temperatures.

The ¹H nuclear magnetic relaxation dispersion (NMRD) profiles were recorded with aqueous solutions of the complexes in the proton Larmor frequency range 0.01–70 MHz, corresponding to magnetic field strengths varying between 2.343×10^{-4} and 1.645 T (Figure 5). The NMRD profiles are typical of small Gd^{3+} chelates and show that the relaxivity of [Gd(1,4-Medo2 ampa)] is higher than that of [Gd(1,7-Medo2 ampa)] in the whole range of proton Larmor frequencies.

The paramagnetic effect of the Gd^{3+} ion on the ¹⁷O chemical shifts of [Gd(1,7-Medo2 ampa)] was found to be negligible. However, the transverse ¹⁷O relaxation rates $1/T_{2r}$ increase with increasing temperature, which is typical of systems in the slow exchange regime (Figure 6). The temperature dependence of the $1/T_{2r}$ data measured for [Gd(1,4-Medo2 ampa)] evidences a fast water-exchange regime in the whole temperature range investigated, although the data display a maximum at the low-temperature side that signals the

commencement of a changeover from a fast water-exchange regime at high temperatures to a slow exchange regime. The temperature dependence of the reduced chemical shifts ($\Delta\omega_r$) measured for [Gd(1,4-Medo2 ampa)] is in line with the $1/T_{2r}$ data.

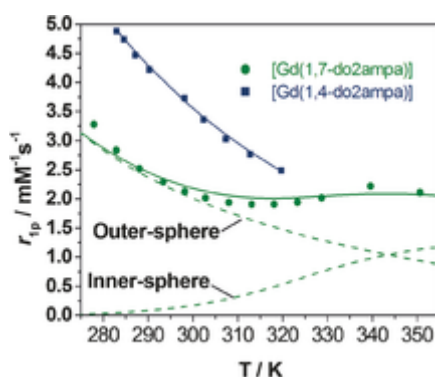


Figure 4. Plot of the ^1H relaxivities (20 MHz, pH 7) of [Gd(1,7-Medo2 ampa)] and [Gd(1,4-Medo2 ampa)] as a function of temperature. The solid lines represent the fits of the data as described in the text. The dotted lines represent the inner- and outer-sphere contributions calculated for [Gd(1,7-Medo2 ampa)].

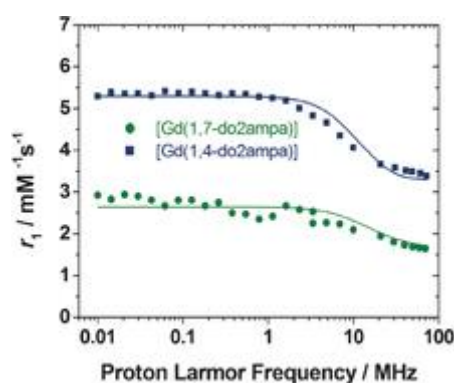


Figure 5. ^1H NMRD profiles recorded for [Gd(1,7-Medo2 ampa)] and [Gd(1,4-Medo2 ampa)] at 298 K. The solid lines represent the fits of the data as described in the text.

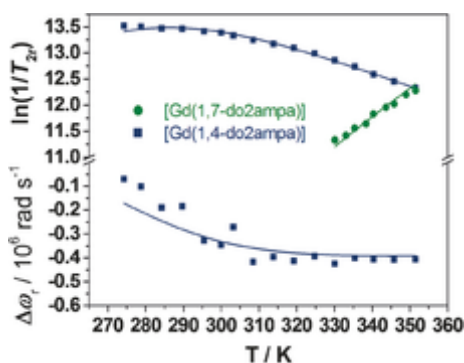
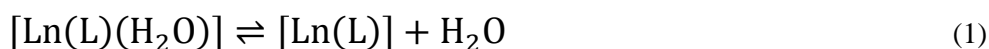


Figure 6. Reduced transverse ^{17}O relaxation rates and chemical shifts measured at 11.74 T (pH 7.2). The solid lines correspond to the fits of the data as described in the text. For [Gd(1,7-Medo2 ampa)] the paramagnetic effect on T_2 was significant only at high temperatures owing to the slow water exchange, whereas effects in chemical shifts were negligible. The reduced relaxation rates and chemical shifts are defined as $1/T_{2r} = [1/(c_{\text{Gd}}/55.5)][1/T_i - 1/T_{iA}]$ and $\Delta\omega_r = [1/(c_{\text{Gd}}/55.5)](\omega - \omega_A)$, in which T_i and T_{2A} are the paramagnetic and diamagnetic relaxation times, ω and ω_A are the paramagnetic and diamagnetic chemical shifts, and c_{Gd} is the concentration of the complex.

A simultaneous analysis of the ^1H relaxivity and ^{17}O NMR data was performed using well-established procedures.^[13] Given the large number of parameters that enter the fit of the NMRD and ^{17}O NMR data, some of them had to be fixed to achieve a reliable analysis. The number of water molecules in the inner coordination sphere of Gd^{3+} was fixed to $q=1$ in $[\text{Gd}(1,7\text{-Medo2 ampa})]$ on the basis of the luminescence measurements described above. However, the ^{17}O $1/T_{2r}$ and chemical shift data obtained for $[\text{Gd}(1,4\text{-Medo2 ampa})]$ clearly point to an equilibrium involving a nine-coordinate species with one inner-sphere water molecule and an eight-coordinate $q=0$ species. This is in line with the hydration numbers obtained from luminescence-lifetime measurements for the complexes with Eu^{3+} and Tb^{3+} ions, which flank Gd^{3+} in the lanthanide series. Hydration equilibria were found to be relatively common in Gd^{3+} complexes. An accurate determination of the equilibrium constant at different temperatures was accomplished in some cases by analyzing the $^5\text{D}_0 \leftarrow ^7\text{F}_0$ transition observed in the absorption spectra of the Eu^{3+} analogues.^[36] These studies provided hydration entropies of $\Delta S^\circ \approx 40 \text{ J mol}^{-1} \text{ K}^{-1}$ for hydration equilibria defined in Equation (1):^[37]



in which L represents a polyaminopolycarboxylate ligand and charges are omitted for simplicity. Thus, we included ΔS° and ΔH° for reaction (1) in $[\text{Gd}(1,4\text{-Medo2 ampa})]$ as fitting parameters. The distance of closest approach for the outer-sphere contribution a_{GdH} was fixed at 4.0 \AA , and the distance between the proton nuclei of the coordinated water molecule and the Gd^{3+} ion (r_{GdH}) was fixed to the values obtained from our DFT calculations (Table 2). The values of the ^{17}O hyperfine coupling constants (A/\hbar , Table 2) were fixed to the values estimated using DFT calculations following the previously reported methodology (TPSSH/SCRECP/EPR-III level).^[29, 38, 39] Finally, the values of the activation energies for the diffusion coefficient (ED_{GdH}), the rotational correlation time (E_r), and the activation energy for the modulation of the zero-field-splitting (E_v) were fixed to common values (22, 20, and 1 kJ mol^{-1} , respectively).^[13] The value of τ_R^{298} of $[\text{Gd}(1,4\text{-Medo2 ampa})]$ was fixed to that obtained for $[\text{Gd}(1,7\text{-Medo2 ampa})]$, a reasonable assumption considering the identical size of the two complexes.

The values obtained for the parameters determining the electron-spin relaxation (the electronic correlation time for the modulation of the zero-field-splitting interaction, τ_v , and the mean square zero-field-splitting energy, Δ^2) and the diffusion coefficient D_{GdH}^{298} are close to those reported for other Gd^{3+} complexes, and the values of the rotational correlation time are consistent with the size of the complexes (Table 2).^[13]

The results of the fit obtained for $[\text{Gd}(1,4\text{-Medo2 ampa})]$ provided a reaction enthalpy of $\Delta H^\circ = 5.7 \pm 3.0 \text{ kJ mol}^{-1}$ for reaction (1) and a reaction entropy of $\Delta S^\circ = 20.6 \pm 10.2 \text{ J mol}^{-1}$, and defined a hydration number of 0.55 at 25°C . This value is in very good agreement with the average of the hydration numbers determined for the Eu^{3+} and Tb^{3+} complexes (Table 1). The hydration number varies from 0.63 at the lowest temperature investigated (1°C) to 0.55 at 78°C . The positive ΔS° value is in line with values obtained for the hydration equilibria of different Eu^{3+} complexes.^[37]

The parameters determined for the water exchange in $[\text{Gd}(1,7\text{-Medo2 ampa})]$ ($k_{\text{ex}}^{298} = 8.8 \pm 1.8 \times 10^3 \text{ s}^{-1}$, $\Delta H^\ddagger = 51.9 \pm 4.4 \text{ kJ mol}^{-1}$) provide a mean residence time of $\tau_m^{298} = 114 \text{ \mu s}$ for the coordinated water molecule at 298 K . This value represents the lowest water-exchange rate of the coordinated water molecule for a Gd^{3+} complex, very similar to the one reported for the SAP isomer of $[\text{Eu}(\text{dotam})(\text{H}_2\text{O})]^{3+}$ (dotam = 2,2',2'',2'''-(1,4,7,10-tetraazacyclododecane-1,4,7,10-tetrayl)tetraacetamide, $k_{\text{ex}}^{298} = 9.4 \times 10^3 \text{ s}^{-1}$),^[40] and somewhat longer than those reported for Gd^{3+} complexes of dota-tetraamides ($9.9\text{--}330 \times 10^3 \text{ s}^{-1}$).^[12] The very low water-exchange rates observed for dota-tetraamide complexes are explained in terms of the rather strong

Gd–water interaction in +3-charged complexes that increases the energy cost for the departure of the water molecule in a dissociative activated exchange mechanism. Thus, the low water-exchange rate in [Gd(1,7-Medo2 ampa)(H₂O)] is surprising considering the neutral charge of the complex. The inner-sphere water molecule in this complex occupies one of the coordination positions in one of the square faces of the square antiprism coordination polyhedron, whereas in dota-like complexes the water molecule is capping that square face. Thus, water molecules occupying the capping position provide weaker Gd–water bonds than water molecules in positions defining the square antiprismatic coordination. As a result, water molecules in the capping position are expected to experience faster water-exchange rates. This is confirmed by the fast water-exchange rate determined for [Gd(1,4-Medo2 ampa)] ($k_{\text{ex}}^{298}=1.2\pm 0.3\times 10^7\text{ s}^{-1}$), which is approximately three times higher than that of [Gd(dota)][−] and very similar to that reported for [Gd(dotma)][−] (Table 2).^[41] Both [Gd(dota)][−] and [Gd(dotma)][−] contain a water molecule coordinated in a capping position. However, the population of [Gd(dotma)][−] in solution is dominated by the TSAP isomer (ca. 81 %), whereas for [Gd(dota)][−] the SAP isomer represents approximately 83 % of the overall population. In both [Gd(dota)][−] and [Gd(dotma)][−] the exchange rate of the coordinated water molecule in the TSAP isomer was reported to be approximately seven times higher than in the SAP form. Thus, the extremely low water-exchange rate of [Gd(1,7-Medo2 ampa)] must be related to the position that the water molecule occupies in the coordination polyhedron. We want to emphasize that the presence of a hydration equilibrium in [Gd(1,4-Medo2 ampa)] introduces some uncertainty on the parameters obtained from the analysis of ¹⁷O NMR and NMRD data. Nevertheless, our data provide clear evidence of a much faster water-exchange rate in [Gd(1,4-Medo2 ampa)] than in [Gd(1,7-Medo2 ampa)].

Table 2. Parameters obtained from the simultaneous analysis of the ¹H relaxivity and ¹⁷O NMR data.

Parameter	1,7-Medo2 ampa	1,4-Medo2 ampa	dota ^{4−[a]}	dotma ^{4−[b]}
q^{298}	1.0	0.55	1.0	1.0
k_{ex}^{298} [10^3 s^{-1}]	8.8±1.8	11 970±3400	4100	11 800
τ_{m}^{298} [μs]	114	0.083	0.24	0.085
ΔH^\ddagger [kJ mol^{-1}]	51.9±4.4	29.8±3.3	49.8	44.5
A/\hbar [10^6 rad s^{-1}]	−3.65 ^[c]	−2.65 ^[c]	−3.7	−3.7
τ_{R}^{298} [ps]	80±27	80 ^[c]	77	81
τ_{V}^{298} [ps]	15.9±2.2	11.4±1.7	11	7.9
Δ^2 [10^{19} s^{-2}]	9.96±1.94	15.7±3.4	1.6	1.7
r_{GdH} [\AA]	2.918 ^[c]	3.068 ^[c]	3.1	3.1
D_{GdH}^{298} [$10^{-10}\text{ m}^2\text{ s}^{-1}$]	21.9±1.3	20.0 ^[b]	20.2	22.4

[a] Data from Ref. [13]; [b] data from Ref. [41]. H₄dotma=α,α',α'',α'''-tetramethyl-1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid; [c] parameters estimated independently using DFT calculations.

The labile capping bond phenomenon is in line with the water-exchange rates measured for a Gd^{3+} complex with a heptadentate triamide cyclen-based ligand. Monodentate binding of phosphate and acetate to this complex, presumably occupying a coordination position in one square face of the polyhedron, was shown to increase the water-exchange rate of the water molecule by two orders of magnitude.^[42] These results also have an implication for the analysis of water-exchange rates of bis-hydrated complexes. For instance, the bis-hydrated complex $[\text{Gd}(\text{do3a})(\text{H}_2\text{O})_2]$ ($\text{H}_3\text{do3a} = 2,2',2''\text{-(1,4,7,10-tetraazacyclododecane-1,4,7-triyl)triacetic acid}$) contains a water molecule coordinating at the apical position and a second water molecule at one of the vertexes of the SAP polyhedron. Previous computational studies suggested that these water molecules should present considerably different exchange rates,^[32] although analysis of ^{17}O NMR data by using a three-site exchange model failed to provide individual exchange rates for the two coordinated water molecules.^[43] Recently, Dolg and co-workers reported computational studies that pointed out that water molecules occupying capping positions within the Ln^{3+} coordination environment are inherently labile.^[44] This phenomenon was attributed to environmental effects because the bond with a ligand occupying a capping position is hindered by the environment. Although these studies provided some hints pointing to the labile capping bond phenomenon, the study presented in this work provides unequivocal experimental evidence for this effect.

Conclusion

A rational design of two ligands with identical donor atoms sets allowed the tuning of the position of the coordinated water molecule, which occupies either a capping position or a position in one of the square faces of the square antiprismatic polyhedron. We found that, whereas $[\text{Gd}(1,7\text{-Medo2 ampa})]$ contains a coordinated water molecule in aqueous solution, the $[\text{Gd}(1,4\text{-Medo2 ampa})]$ analogue presents an equilibrium in solution involving $q=1$ and $q=0$ species. Nevertheless, the water-exchange rate measured for the fraction of complex containing a coordinated water molecule is three orders of magnitude higher than that of $[\text{Gd}(1,7\text{-Medo2 ampa})]$. Thus, the present contribution provides solid evidence demonstrating that water ligands occupying capping positions in the coordination polyhedron are intrinsically labile. The labile capping bond phenomenon demonstrated here represents a significant advance for the rational design of optimized Gd^{3+} -based contrast agents and CEST probes for MRI applications in clinical diagnostics and pre-clinical research. Furthermore, it is likely that this effect occurs not only in Ln^{3+} complexes but also in complexes with other metal ions, paving the way to a more rational control of the reactivity of metal complexes.

Acknowledgements

A.R.R., M.R.F., D.E.G., T.R.B., and C.P.I. thank Ministerio de Economía y Competitividad (CTQ2009-10721/PPQ, CTQ2013-43243-P, and CTQ2015-71211-REDT) and Xunta de Galicia (CN2012/011) for generous financial support. M.B. and F.C. are grateful to Università del Piemonte Orientale for a research grant. R.T. and V.P. thank the University of Brest and the CG29 for a post-doctoral research grant (A.R.R.). The research presented in this work was supported by the EU and co-financed by the European Regional Development Fund under the project GINOP-2.3.2.-15-2016-00008 and by the Hungarian Scientific Research Fund (OTKA K-109029 and K-120224 projects). This work was also supported by the János Bolyai Research Scholarship (G.T.) of the Hungarian Academy of Sciences. The authors are indebted to Centro de Supercomputación of Galicia (CESGA) for providing the computer facilities.

- [1] L. Helm, A. E. Merbach, *Chem. Rev.* 2005, 105, 1923–1959.
- [2] a) N. P. E. Barry, P. J. Sadler, *Pure Appl. Chem.* 2014, 86, 1897–1910; b) D. T. Richens, *Chem. Rev.* 2005, 105, 1961–2002; c) E. Balogh, W. H. Casey, *Prog. Nucl. Magn. Reson. Spectrosc.* 2008, 53, 193–207.
- [3] a) J. Maigut, R. Meier, A. Zahl, R. van Eldik, *J. Am. Chem. Soc.* 2008, 130, 14556–14569; b) A. Brausam, J. Maigut, R. Meier, P. A. Szilagy, H.-J. Buschmann, W. Massa, Z. Homonnay, R. van Eldik, *Inorg. Chem.* 2009, 48, 7864–7884; c) R. van Eldik, *Coord. Chem. Rev.* 2007, 251, 1649–1662.
- [4] A. Cusanelli, U. Frey, D. T. Richens, A. E. Merbach, *J. Am. Chem. Soc.* 1996, 118, 5265–5271.
- [5] P. Caravan, E. Toth, A. Rothenbauer, A. E. Merbach, *J. Am. Chem. Soc.* 1999, 121, 10403–10409.
- [6] a) C. Cossy, L. Helm, A. E. Merbach, *Inorg. Chem.* 1988, 27, 1973–1979; b) C. Cossy, L. Helm, A. E. Merbach, *Inorg. Chem.* 1989, 28, 2699–2703.
- [7] a) *The Chemistry of Contrast Agents in Medical Magnetic Resonance Imaging 2nd Ed.*, (Eds.: A. E. Merbach, L. Helm, É. Tóth), Wiley, Hoboken, 2013; b) M. C. Heffern, L. M. Matosziuk, T. J. Meade, *Chem. Rev.* 2014, 114, 4496–4539.
- [8] a) P. Caravan, J. J. Ellison, T. J. McMurry, R. B. Lauffer, *Chem. Rev.* 1999, 99, 2293–2352; b) Y. Song, E. K. Kohlmeier, T. J. Meade, *J. Am. Chem. Soc.* 2008, 130, 6662–6663.
- [9] a) P. Caravan, *Chem. Soc. Rev.* 2006, 35, 512–523; b) S. Aime, M. Botta, M. Fasano, E. Terreno, *Acc. Chem. Res.* 1999, 32, 941–949; c) B. N. Siriwardena-Mahanama, M. J. Allen, *Molecules* 2013, 18, 9352–9381; d) A. T. Preslar, G. Parigi, M. T. McClendon, S. S. Sefick, T. J. Moyer, C. R. Haney, E. A. Waters, K. W. MacRenaris, C. Luchinat, S. I. Stupp, T. J. Meade, *ACS Nano* 2014, 8, 7325–7332.
- [10] a) S. Aime, S. Geninatti Crich, E. Gianolio, G. B. Giovenzana, L. Tei, E. Terreno, *Coord. Chem. Rev.* 2006, 250, 1562–1579; b) E. Terreno, D. Delli Castelli, S. Aime, *Contrast Media Mol. Imaging* 2010, 5, 78–98.
- [11] a) E. Vinogradov, A. D. Sherry, R. E. Lenkinski, *J. Magn. Reson.* 2013, 229, 155–172; b) P. C. M. van Zijl, N. N. Yadav, *Magn. Reson. Med.* 2011, 65, 927–948; c) X. Yang, N. N. Yadav, X. Song, S. R. Banerjee, H. Edelman, I. Minn, P. C. M. van Zijl, M. G. Pomper, M. T. McMahon, *Chem. Eur. J.* 2014, 20, 15824–15832.
- [12] S. Aime, A. Barge, J. I. Bruice, M. Botta, J. A. K. Howard, J. M. Moloney, D. Parker, A. S. de Sousa, M. Woods, *J. Am. Chem. Soc.* 1999, 121, 5762–5771.
- [13] D. H. Powell, O. M. Ni Dhubghaill, D. Pubanz, L. Helm, Y. S. Lebedev, W. Schlaepfer, A. E. Merbach, *J. Am. Chem. Soc.* 1996, 118, 9333–9346.
- [14] M. Mato-Iglesias, C. Platas-Iglesias, K. Djanashvili, J. A. Peters, E. Toth, E. Balogh, R. N. Muller, L. Vander Elst, A. de Blas, T. Rodriguez-Blas, *Chem. Commun.* 2005, 4729–4731.
- [15] a) F. A. Dunand, S. Aime, A. E. Merbach, *J. Am. Chem. Soc.* 2000, 122, 1506–1512; b) W. T. Dixon, J. Ren, A. J. M. Lubag, J. Ratnakar, E. Vinogradov, I. Hancu, R. E. Lenkinski, A. D. Sherry, *Magn. Reson. Med.* 2010, 63, 625–632.
- [16] a) W. S. Fernando, A. F. Martins, P. Zhao, Y. Wu, G. E. Kiefer, C. Platas-Iglesias, A. D. Sherry, *Inorg. Chem.* 2016, 55, 3007–3014; b) N. Cakic, T. Savic, J. Stricker-Shaver, V. Truffault, C. Platas-Iglesias, C. Mirkes, R. Pohmann, K. Scheffler, G. Angelovski, *Chem. Commun.* 2016, 52, 9224–9227.

- [17] S. Laurent, L. Vander Elst, F. Botteman, R. N. Muller, *Eur. J. Inorg. Chem.* 2008, 4369–4379.
- [18] a) R. Ruloff, E. Toth, R. Scopelliti, R. Tripier, H. Handel, A. E. Merbach, *Chem. Commun.* 2002, 2630–2631; b) S. Laus, R. Ruloff, E. Toth, A. E. Merbach, *Chem. Eur. J.* 2003, 9, 3555–3566; c) J. Kotek, P. Lebduskova, P. Hermann, L. Vander Elst, R. N. Muller, C. F. G. C. Geraldès, T. Maschmeyer, I. Lukes, J. A. Peters, *Chem. Eur. J.* 2003, 9, 5899–5915; d) A. Congreve, D. Parker, E. Gianolio, M. Botta, *Dalton Trans.* 2004, 1441–1445.
- [19] A. L. Thompson, D. Parker, D. A. Fulton, J. A. K. Howard, S. U. Pandya, H. Puschmann, K. Senanayake, P. A. Stenson, A. Badari, M. Botta, S. Avedano, S. Aime, *Dalton Trans.* 2006, 5605–5616.
- [20] M. Regueiro-Figueroa, B. Bensenane, E. Ruscsak, D. Esteban-Gómez, L. J. Charbonniere, G. Tircso, I. Toth, A. de Blas, T. Rodriguez-Blas, C. Platas-Iglesias, *Inorg. Chem.* 2011, 50, 4125–4141.
- [21] M. Le Baccon, F. Chuburu, L. Toupet, H. Handel, M. Soibinet, I. Dechamps-Olivier, J.-P. Barbier, Aplincourt, *New J. Chem.* 2001, 25, 1168–1174.
- [22] J. Rohovec, R. Gyepes, I. Cisarova, J. Rudovsky, I. Lukes, *Tetrahedron Lett.* 2000, 41, 1249–1253.
- [23] M. Mato-Iglesias, A. Roca-Sabio, Z. Palinkas, D. Esteban-Gomez, C. Platas-Iglesias, E. Toth, A. de Blas, T. Rodriguez-Blas, *Inorg. Chem.* 2008, 47, 7840–7851.
- [24] a) C. Li, W.-T. Wong, *J. Org. Chem.* 2003, 68, 2956–2959; b) W. J. Kruper Jr. , P. R. Rudolf, C. A. Langhoff, *J. Org. Chem.* 1993, 58, 3869–3876.
- [25] a) L. S. Natrajan, A. J. L. Villaraza, A. M. Kenwright, S. Faulkner, *Chem. Commun.* 2009, 6020–6022; b) M. Main, M. M. Meloni, M. Jauregui, D. Sykes, S. Faulkner, A. M. Kenwright, J. S. Snaith, *Chem. Commun.* 2008, 5212–5214.
- [26] a) W. D. W. Horrocks, Jr. , D. R. Sudnick, *J. Am. Chem. Soc.* 1979, 101, 334–340; b) R. M. Supkowski, W. D. W. Horrocks Jr. , *Inorg. Chim. Acta* 2002, 340, 44–48.
- [27] A. Beeby, I. M. Clarkson, R. S. Dickins, S. Faulkner, D. Parker, L. Royle, A. S. de Sousa, J. A. G. Williams, M. Woods, *J. Chem. Soc. Perkin Trans. 2* 1999, 493–503.
- [28] M. Dolg, H. Stoll, A. Savin, H. Preuss, *Theor. Chim. Acta* 1989, 75, 173–194.
- [29] J. Tao, J. P. Perdew, V. N. Staroverov, G. E. Scuseria, *Phys. Rev. Lett.* 2003, 91, 146401–146404.
- [30] Gaussian 09, Revision D.01, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery, Jr. , J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski, D. J. Fox, Gaussian, Inc. Wallingford CT, 2009.
- [31] D. Esteban-Gómez, A. de Blas, T. Rodriguez-Blas, L. Helm, C. Platas-Iglesias, *ChemPhysChem* 2012, 13, 3640–3650.
- [32] M. Regueiro-Figueroa, C. Platas-Iglesias, *J. Phys. Chem. A* 2015, 119, 6436–6445.

- [33] J. A. Peters, *Contrast Media Mol. Imaging* 2016, 11, 160–168.
- [34] S. Aime, A. Barge, M. Botta, D. Parker, A. S. De Sousa, *J. Am. Chem. Soc.* 1997, 119, 4767–4768.
- [35] C. Platas, F. Avecilla, A. de Blas, T. Rodríguez-Blas, C. F. G. C. Geraldès, E. Toth, A. E. Merbach, J.-C. G. Bünzli, *J. Chem. Soc. Dalton Trans.* 2000, 611–618.
- [36] N. Graepi, D. H. Powell, G. Laurency, L. Zekany, A. E. Merbach, *Inorg. Chim. Acta* 1995, 235, 311–326.
- [37] a) C. Platas-Iglesias, D. M. Corsi, L. Vander Elst, R. N. Muller, D. Imbert, J.-C. G. Bünzli, E. Toth, T. Maschmeyer, J. A. Peters, *Dalton Trans.* 2003, 727–737; b) E. Balogh, M. Mato-Iglesias, C. Platas-Iglesias, E. Toth, K. Djanashvili, J. A. Peters, A. de Blas, T. Rodríguez-Blas, *Inorg. Chem.* 2006, 45, 8719–8728.
- [38] M. Dolg, H. Stoll, H. Preuss, *J. Chem. Phys.* 1989, 90, 1730–1734.
- [39] N. Rega, M. Cossi, V. Barone, *J. Chem. Phys.* 1996, 105, 11060–11067.
- [40] F. A. Dunand, R. S. Dickins, D. Parker, A. E. Merbach, *Chem. Eur. J.* 2001, 7, 5160–5167.
- [41] S. Aime, M. Botta, Z. Garda, B. E. Kucera, G. Tircso, V. G. Young, M. Woods, *Inorg. Chem.* 2011, 50, 7955–7965.
- [42] J. I. Bruce, R. S. Dickins, L. J. Govenlock, T. Gunnlaugsson, S. Lopinski, M. P. Lowe, D. Parker, R. D. Peacock, J. J. B. Perry, S. Aime, M. Botta, *J. Am. Chem. Soc.* 2000, 122, 9674–9684.
- [43] E. Tóth, O. M. N. Dhubhghaill, G. Besson, L. Helm, A. E. Merbach, *Magn. Reson. Chem.* 1999, 37, 701–708.
- [44] a) J. Zhang, N. Heinz, M. Dolg, *Inorg. Chem.* 2014, 53, 7700–7708; b) J. Zhang, M. Dolg, *J. Phys. Chem. A* 2015, 119, 774–780.