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Paramagnetic lanthanide chelates for multicontrast MRI†

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The preparation of a paramagnetic chelator that serves as a platform for multicontrast MRI, and can be utilized either as a T_1 -weighted, paraCEST or ^{19}F MRI contrast agent is reported. Its europium(III) complex exhibits an extremely slow water exchange rate which is optimal for the use in CEST MRI. The potential of this platform was demonstrated through a series of MRI studies on tube phantoms and animals.

Visualization of various biological processes that take place at cellular and molecular levels is the main goal of modern diagnostic and molecular imaging techniques. Due to different sensitivity, penetration depth, spatial or temporal resolution properties of the available imaging methods, the development of multimodal imaging experienced great advancements in the last decade.^{1,2} Nevertheless, a combination of two (or more) techniques is often challenging, requiring integration of different physical phenomena in the common hardware, or careful design of multimodal imaging probes.^{3,4}

Today, magnetic resonance imaging (MRI) is one of the most powerful imaging tools capable of displaying an excellent soft tissue contrast and furthermore different types of contrasts. A number of diverse MRI contrast agents can improve the specificity of MRI measurements and, based on their nature, can provide different types of information. The most commonly used ^1H -MRI contrast agents are paramagnetic Gd^{3+} -based complexes or superparamagnetic iron-oxide based nanoparticles (T_1 - and T_2 -shortening

agents, respectively).⁵ Recently, an entirely different mechanism for altering an MRI contrast, based on chemical exchange saturation transfer (CEST), has been developed. Contrast agents for CEST imaging usually consist of paramagnetic chelates (paraCEST agents) specifically designed to shift the resonances of exchangeable protons (NH, OH or bound water) further away from the bulk water.⁶ One of the great advantages of CEST is the detectability of its effect in combination with the presaturation RF pulse; only when this RF is applied at the specific frequency of the exchangeable protons, the MRI contrast will appear. Consequently, this method allows the multi-frequency readout and adjustment of the frequencies (e.g. by choice of the group with exchangeable protons or choice of the paramagnetic ion), which can be used for separate detection and visualization of different cellular and tissue environments.^{7,8} Finally, MRI can be performed on heteronuclear and hyperpolarized systems.⁹ Here, the best choice is ^{19}F NMR because of its high sensitivity, the easy re-tuning of standard MRI instruments from ^1H to ^{19}F nuclei, the high natural abundance of the ^{19}F isotope and the absence of background signals from intrinsic biomolecules, thus allowing quantitative studies.^{10,11}

With such potential to provide diverse information by employing different frequencies and contrast mechanisms, MRI could be exploited for concurrent studies by means of T_1 -weighted, CEST and ^{19}F imaging protocols. To this aim, it is highly desirable to develop multicontrast agents, probes capable of making use of these different types of MR contrast, thus resulting in a set of unique information related to the studied tissue of interest.

We therefore aimed to prepare an agent that is capable of chelating the paramagnetic ion, possesses frequency-shifted protons in slow exchange with bulk water, and finally bears a sufficient number of fluorine atoms to provide a sizeable ^{19}F NMR signal (Fig. 1). We designed ligand **L**, a derivative of DOTA-tetra-glycineamide (DOTAM-Gly),¹² which displays CEST properties and ensures good solubility in aqueous solutions, especially after incorporation of fluorine atoms. The fluorinated moieties were introduced in the molecule at *trans* positions of the macrocyclic ring through two CF_3 groups by using a 4-(trifluoromethyl)-*L*-phenylalanine (*p*- CF_3 -Phe) derivative and a convenient six-step

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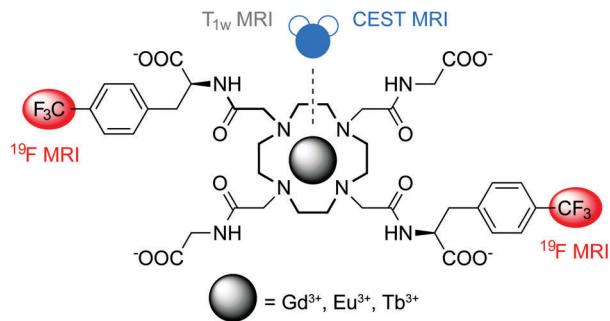


Fig. 1 The chemical structure of chelator **L** described in this work: paramagnetic ion makes it suitable for T_1 -weighted or paraCEST MRI, while nearby CF_3 groups enable ^{19}F NMR, concurrently taking advantage of the paramagnetic enhancement effect.

synthetic procedure (ESI †). The resulting molecule **L** underwent complex formation with the Gd^{3+} , Eu^{3+} and Tb^{3+} ions and the properties of **GdL**, **EuL** and **TbL** were further investigated (ESI †). Their overall charge should be advantageous (negative) due to the presence of four carboxylate groups, while appropriate complex stability necessary for *in vivo* experiments is ensured by the kinetic inertness provided by the tetraamide moieties.¹³ Finally, rigid benzyl moieties may establish, compared to *e.g.* aliphatic spacers, a well-defined distance of ^{19}F nuclei from the paramagnetic center, thus providing advantageous relaxation properties at the frequency of these nuclei.¹⁴

The longitudinal relaxivity of **GdL** was calculated by obtaining T_1 values of a series of buffered solutions at different complex concentrations. The resulting value of $3.1 \text{ mM}^{-1} \text{ s}^{-1}$ is expected for monohydrated tetraamide complexes with a slow water exchange rate.¹⁵ CEST measurements were performed by selective presaturation of the **EuL** and **TbL** samples in incremental steps over a range of frequencies followed by plotting the remaining steady-state bulk water signal, M_z/M_0 , as a function of saturation frequency (Fig. 2). The resulting Z-spectra of **EuL** exhibited a strong CEST effect ($\sim 50\%$ at 25°C , 15 mM , $B_1 = 25.0 \mu\text{T}$) centred at 50 ppm relative to bulk water, which is commonly associated with the water molecule directly coordinated to Eu^{3+} . The CEST effect remains strong at 37°C along with slight broadening and shifting of the CEST signal (5 ppm) towards the bulk water resonance due to the hyperfine shift effect of Eu^{3+} (Fig. 2, top).¹⁶

Water exchange rates at both temperatures were determined using a concentration-independent method previously developed by Shery and colleagues,¹⁶ and were confirmed by quantification of the exchange rates as a function of saturation time or saturation power (QUEST and QUESP experiments, respectively).¹⁷ The values obtained from these experiments revealed extremely slow exchange rates of around 1 and 2 kHz or bound-water lifetimes (τ_M) of around 1 ms and $500 \mu\text{s}$ at 25 and 37°C , respectively (Table 1). These residence times are as long as those reported very recently for the phosphonate esters of DOTAM-Gly at 25°C ,¹⁸ likely due to the presence of hydrophobic *p*- CF_3 -Phe moieties. Consequently, **EuL** displays an almost optimal τ_M which lies in the range of 10^{-4} – 10^{-2} s , allowing CEST experiments using weaker B_1 fields. On the other hand, **TbL** displayed a weak CEST effect for both the inner-sphere water molecule and amide

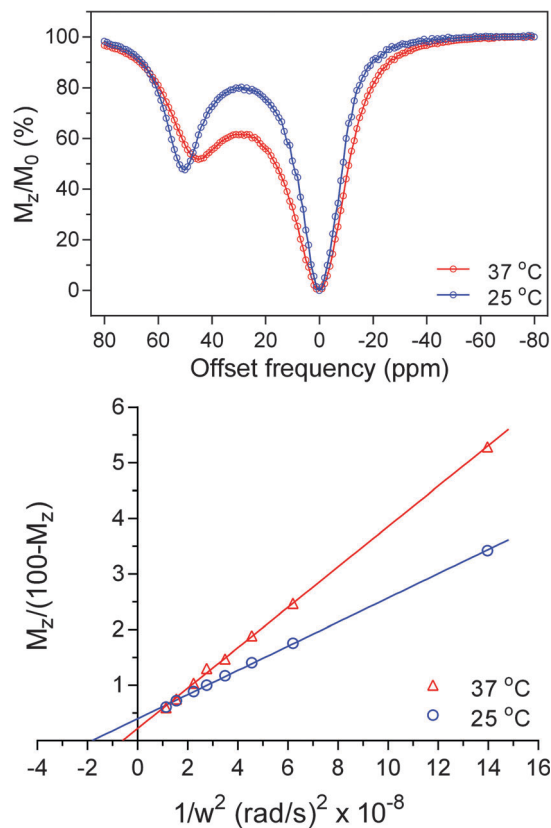


Fig. 2 CEST experiments with **EuL** (15 mM). (top) Z-spectra of **EuL** at $B_1 = 25.0 \mu\text{T}$ and irradiation time = 5 s ; (bottom) determination of water exchange rates at 25°C (blue) and 37°C (red) at $B_1 = 35.0, 30.0, 25.0, 22.5, 20.0, 17.5, 15.0$ and $10.0 \mu\text{T}$, and irradiation time = 10 s .

Table 1 Water exchange rates (in Hz) obtained for **EuL** using the two independent methods

Temperature	Omega plots	QUEST/QUESP
25°C	1176 ± 22	930 ± 11
37°C	2036 ± 110	2000 ± 138

protons (ESI †), probably for the same reasons related to slow water exchange as in **EuL**.^{19,20}

^{19}F NMR spectra of **GdL**, **EuL** and **TbL** were recorded and ^{19}F relaxation rates were determined to estimate the potential of these agents for ^{19}F MRI. **GdL** displayed a single broad ^{19}F NMR resonance, indicating substantial shortening of the relaxation times, while **EuL** and **TbL** revealed the existence of at least three paramagnetic species in the solution with different ^{19}F chemical shifts (ESI †). While the two peaks with higher intensities can be assigned to the common monocapped square-antiprismatic and monocapped twisted square-antiprismatic (SAP and TSAP, respectively) isomers of this type of compound,²¹ the appearance of the third resonance with very weak intensity can be explained by racemization of the starting amino acid or racemization of alkylating arms prior to or during alkylation of cyclen, as previously observed for similar systems under comparable experimental conditions.²² Consequently, ^{19}F relaxation rates were determined only for the most abundant peaks (Table 2).



Table 2 ^{19}F relaxation rates of **GdL**, **EuL** and **TbL** (300 MHz, 25 °C)

Temperature	^{19}F R_1 (Hz)	^{19}F R_2 (Hz)
GdL	182	385
EuL	1.4 ^{a,b}	18 ^a 19 ^b
TbL	32 ^a 12 ^b	48 ^a 28 ^b

^a More abundant isomer. ^b Less abundant isomer.

As expected, **GdL** dramatically affected both ^{19}F R_1 and R_2 , while still keeping a favorable R_1/R_2 ratio to obtain a good signal-to-noise ratio (SNR), when using ultrafast sequences.²³ **EuL** and **TbL** also enhanced ^{19}F R_1 and R_2 , however the reduction in the R_1/R_2 ratio (in the case of **EuL**) or larger signal splitting (in the case of **TbL**) suggests **GdL** as a better candidate for ^{19}F MRI studies, with the current DOTA-type chelator.

The potential of these systems to serve as multicontrast agents was demonstrated using *in vitro* MRI on tube phantoms (Fig. 3a–d). Four tubes containing **GdL**, **EuL**, **TbL** and water as controls were imaged using a 7T MRI scanner by different imaging protocols and frequencies. The greatest effect on relaxation times and hence the signal enhancement in T_1 -weighted MRI experiments was observed for **GdL** (Fig. 3a), as would be expected; this effect was also confirmed by T_2 -weighted MRI experiments (Fig. 3b). All three complexes displayed very good ^{19}F MRI contrasts using the sequence parameters adjusted to assume different ^{19}F relaxation rates due to influence of different paramagnetic ions (Fig. 3c). Lastly, only **EuL** exhibited a strong contrast in the CEST MRI experiment (Fig. 3d), as already indicated above in the NMR CEST experiments.

To confirm this platform as a multicontrast MRI agent in a complex environment, we have carried out an MRI study on animals using **GdL** and **EuL**. The contrast agent was injected intracranially into the somatosensory cortex of the anesthetized rats outside the scanner. For **GdL**, the animal was transferred into the scanner, and a very strong T_1 -weighted MRI signal was recorded *in vivo* (Fig. 3e). Slow reduction in the MRI signal over a period of a few hours indicated very slow diffusion of **GdL** and its potential interaction with brain tissues. Although a similar behavior was previously observed with aminobisphosphonate-containing contrast agents,²⁴ it is hard to rationalize this effect with the current chelating platform since its diffusion properties in solution did not indicate any aggregation, *i.e.* the diffusion coefficient corresponded to other monomeric agents of similar size (ESI^+).

Additionally, the effect of **EuL** was assessed by means of MRI *ex vivo*. A weak T_1 -weighted MRI contrast was obtained as expected after the animal was euthanized and transferred into the scanner (Fig. 3f). In parallel, a strong CEST contrast ($\sim 10\%$ signal change) was successfully obtained at the frequency of the inner-sphere water molecule bound to Eu^{3+} (Fig. 3g), confirming the great potential of **EuL** for further paraCEST studies due to its long τ_M (*vide supra*). In regard to ^{19}F MRI, several imaging sequences were tested on both **GdL** and **EuL**, using non-fluorine containing anesthesia to avoid possible interferences at the fluorine frequency. However, only a slight change in signal intensity could

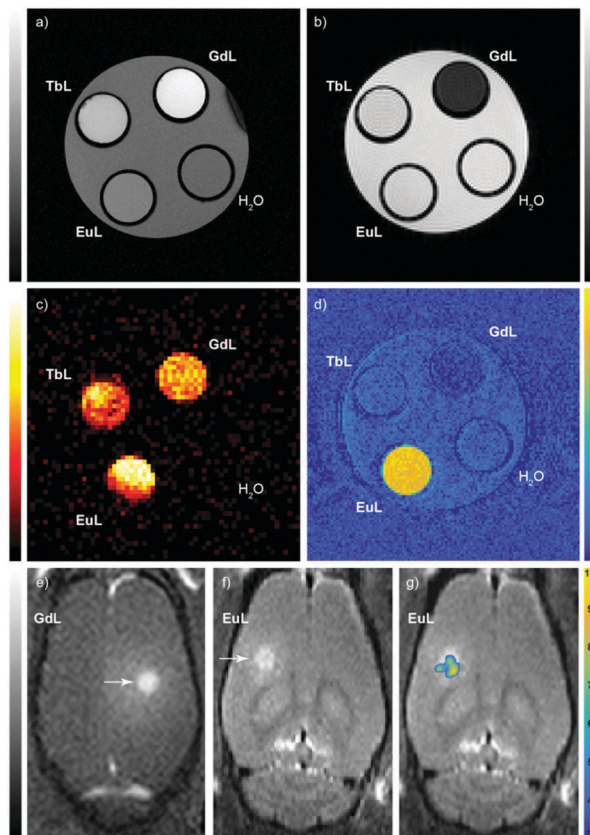


Fig. 3 MRI experiments with the multicontrast agent. *In vitro* MRI on tube phantoms with 5 mM of complexes (pH 7.3, HEPES, 25 °C): (a) T_1 -weighted MRI; (b) T_2 -weighted MRI; (c) ^{19}F MRI; (d) paraCEST MRI. *In vivo* (**GdL**) and *ex vivo* (**EuL**) MRI in rat cortex: (e) T_1 -weighted MRI with **GdL**; (f) T_1 -weighted MRI with **EuL**; (g) merged CEST and T_1 -weighted MRI with **EuL**. Arrows indicate the region where the contrast agent was injected; bars show signal intensity in arbitrary units except %CEST in (g).

be detected after 2.3 hours of monitoring. As already discussed above, the lack of signals could indicate an interaction of the agent with surrounding tissues, which significantly reduces ^{19}F T_2 relaxation times and leads to signal disappearance (a notable broadening of the ^{19}F signal in the measured 1D spectrum was observed for **GdL**). However, this effect can likely be avoided with another experimental design where the tissue density is lower (*e.g.* in the blood stream or kidneys), or by combining this platform to a nanosized system that will prevent any interaction with the tissue and improve the agent's biocompatibility.²⁵

In conclusion, we report a promising platform for the development of multicontrast agents for MRI. A small size molecule accommodates different paramagnetic ions and subsequently enhances ^1H T_1 -weighted, ^1H CEST, and ^{19}F MRI contrasts. The **GdL** and **EuL** complexes can concurrently serve as ^1H T_1 -weighted and ^{19}F MRI, or as ^1H CEST and ^{19}F MRI agents, respectively. As these two complexes are expected to have essentially the same biodistribution, the three different contrast mechanisms could be exploited using the same molecular platform, even if **GdL** and **EuL** have to be administered separately.²⁶ Furthermore, the installation of the aromatic fluorinated units likely caused extremely advantageous exchange rates in **EuL** for CEST experiments,²⁰



while possibly deteriorating the agent's biocompatibility for ^{19}F MRI. Nevertheless, further improvements should easily be envisaged to provide the optimized multicontrast agent. Structural optimizations towards adjustment of the chelator's coordination properties may lead to a single isomer species that will be beneficial for ^{19}F MRI. Synthetic modifications can lead to a higher number of fluorine atoms per molecule to increase the signal intensity, while a combination with various nanocarriers can improve the delivery, biokinetics and potentiality also the T_2 contrast of this multicontrast agent. The ability to collect different types of information from a single imaging probe just by using different imaging protocols brings new quality to MRI and can be a great asset for current molecular imaging to study various biological phenomena.

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The animal experiments were approved by the local authorities (Regierungspraesidium), and were in compliance with the guidelines of the European directive (2010/63/EU) for the care and protection of animals used for scientific purposes.

Notes and references

1 R. Weissleder and M. J. Pittet, *Nature*, 2008, **452**, 580–589.

2 H. Kobayashi, M. R. Longmire, M. Ogawa and P. L. Choyke, *Chem. Soc. Rev.*, 2011, **40**, 4626–4648.

3 S. R. Cherry, *Annu. Rev. Biomed. Eng.*, 2006, **8**, 35–62.

4 A. Y. Louie, *Chem. Rev.*, 2010, **110**, 3146–3195.

5 P. Caravan, in *Molecular and cellular MR imaging*, ed. M. M. J. Modo and J. W. M. Bulte, CRC Press, Taylor & Francis Group, Boca Raton, London, New York, 2007, pp. 13–36.

6 A. D. Sherry and M. Woods, *Annu. Rev. Biomed. Eng.*, 2008, **10**, 391–411.

7 S. Aime, C. Carrera, D. D. Castelli, S. G. Crich and E. Terreno, *Angew. Chem., Int. Ed.*, 2005, **44**, 1813–1815.

8 S. Viswanathan, S. J. Ratnakar, K. N. Green, Z. Kovacs, L. M. De Leon-Rodríguez and A. D. Sherry, *Angew. Chem., Int. Ed.*, 2009, **48**, 9330–9333.

9 E. Terreno, D. D. Castelli, A. Viale and S. Aime, *Chem. Rev.*, 2010, **110**, 3019–3042.

10 I. Tirrotta, V. Dichiarante, C. Pigliacelli, G. Cavallo, G. Terraneo, F. B. Bombelli, P. Metrangolo and G. Resnati, *Chem. Rev.*, 2015, **115**, 1106–1129.

11 M. Srinivas, A. Heerschap, E. T. Ahrens, C. G. Figdor and I. J. M. d. Vries, *Trends Biotechnol.*, 2010, **28**, 363–370.

12 S. Aime, A. Barge, D. Delli Castelli, F. Fedeli, A. Mortillaro, F. U. Nielsen and E. Terreno, *Magn. Reson. Med.*, 2002, **47**, 639–648.

13 E. Brücher, G. Tircsó, Z. Baranyai, Z. Kovács and A. D. Sherry, *The Chemistry of Contrast Agents in Medical Magnetic Resonance Imaging*, John Wiley & Sons, Ltd, 2013, pp. 157–208.

14 P. Harvey, I. Kuprov and D. Parker, *Eur. J. Inorg. Chem.*, 2012, 2015–2022.

15 S. Aime, A. Barge, J. I. Bruce, M. Botta, J. A. K. Howard, J. M. Moloney, D. Parker, A. S. de Sousa and M. Woods, *J. Am. Chem. Soc.*, 1999, **121**, 5762–5771.

16 W. T. Dixon, J. M. Ren, A. J. M. Lubag, J. Ratnakar, E. Vinogradov, I. Hancu, R. E. Lenkinski and A. D. Sherry, *Magn. Reson. Med.*, 2010, **63**, 625–632.

17 M. T. McMahon, A. A. Gilad, J. Y. Zhou, P. Z. Sun, J. W. M. Bulte and P. C. M. van Zijl, *Magn. Reson. Med.*, 2006, **55**, 836–847.

18 W. S. Fernando, A. F. Martins, P. Zhao, Y. Wu, G. E. Kiefer, C. Platas-Iglesias and A. D. Sherry, *Inorg. Chem.*, 2016, **55**, 3007–3014.

19 X. J. Wang, Y. K. Wu, T. C. Soesbe, J. Yu, P. Y. Zhao, G. E. Kiefer and A. D. Sherry, *Angew. Chem., Int. Ed.*, 2015, **54**, 8662–8664.

20 A. D. Sherry and Y. K. Wu, *Curr. Opin. Chem. Biol.*, 2013, **17**, 167–174.

21 J. A. Peters, K. Djanashvili, C. F. G. C. Geraldès and C. Platas-Iglesias, *The Chemistry of Contrast Agents in Medical Magnetic Resonance Imaging*, John Wiley & Sons, Ltd, 2013, pp. 209–276.

22 T. Mani, A. C. L. Opina, P. Y. Zhao, O. M. Evbuomwan, N. Milburn, G. Tircsó, C. Kumas and A. D. Sherry, *JBIC, J. Biol. Inorg. Chem.*, 2014, **19**, 161–171.

23 F. Schmid, C. Höltske, D. Parker and C. Faber, *Magn. Reson. Med.*, 2013, **69**, 1056–1062.

24 I. Mamedov, S. Canals, J. Henig, M. Beyerlein, Y. Murayama, H. A. Mayer, N. K. Logothetis and G. Angelovski, *ACS Chem. Neurosci.*, 2010, **1**, 819–828.

25 K. J. Thurecht, I. Blakey, H. Peng, O. Squires, S. Hsu, C. Alexander and A. K. Whittaker, *J. Am. Chem. Soc.*, 2010, **132**, 5336–5337.

26 A. M. Funk, V. Clavijo Jordan, A. D. Sherry, S. J. Ratnakar and Z. Kovacs, *Angew. Chem., Int. Ed.*, 2016, **55**, 5024–5027.

